Green Revolution – genes in modern crop varieties

Introduction

Global crop grain production grew from 1 billion tons in 1960 to 2 billion tons by 2000 (Khush, 2001). This growth was a result of a series of plant breeding and agronomy research initiatives, known as the Green Revolution, led by Norman Borlaug, who received the Nobel Peace Prize in 1970 for being the "Father of the Green Revolution". A part of these activities was an innovative wheat research program, and the foundation of the International Maize and Wheat Improvement Centre (CIMMYT) based on Borlaug's pioneering work in Mexico. The motivation for this program was the increasing pressure of food-population imbalance which was expected to lead to worldwide famines.

During the 1940s years, tall wheat varieties (e.g. Mentana) (Fisher and Stapper, 1987) were grown and the average wheat yield in Mexico was 2 t/ha. From 1900 to 1949, height of the plants ranged from 120.9 cm to 113.7 cm (Würschum et al., 2017). These were susceptible to lodging which led to loss of grains and therefore, yield. The harvest index, calculated from the yield of a crop species versus the total amount of biomass, was 0.3 (Kush, 2001). This meant that wheat plants produced 30% grain and 70% straw and their yield potential was 4 t/ha. The addition of fertiliser caused the plants to grow even taller, were more prone to lodging and tillering. The crops could not fully exploit the additional nutrients and the maximum yield was predicted not being able to cope with the growing population.

Wheat stem rust (*Puccinia graminis*) infection was spreading in Mexico. Borlaug noticed that new wheat varieties could be grown twice in a year in between the Ciudad Obregon and Toluca field sites (Figure 1) (Ortiz et al., 2007). This meant the varieties were grown from spring to October at Toluca (2,500m in altitude), and 'shuttled' to Ciudad Obregon from November. This 'shuttle breeding' led to more resistant wheat varieties as they were exposed to different environments and diseases and sped up the breeding process. CIMMYT defined 12 mega-environments and the breeding would take place at two different sites and the varieties' yield would be compared in the different environments. This was the beginning of the Green Revolution.

The major breakthrough was the introduction of a Japanese cross between Daruma with an American variety which led to Norin 10 (Hedden, 2003). It was Norin10-Brevor 14, produced by Norman Borlaug, which had the Norin 10 dwarfing gene and was suitable for temperate climate. The introduction of the dwarfing gene increased the harvest index to 0.5 by shifting the biomass production to more grains, rather than straw production (Khush, 1993). Shorter plant height led also enabled the optimisation of fertiliser usage by the crops without lodging or tillering. Their maximum yield potential



Figure 1. Shuttle breeding started in Mexico (left) to enhance wheat rust resistance and international shuttle breeding has led to even more resistant modern varieties (Ortiz et al., 2007). The introduction of Norin 10 dwarfing genes has reduced the height of wheat plants (right).

increased from 4 t/ha to 10 t/ha which enabled the food production to more than double from 1966 to 1990 (Kush, 1999).

Approximately 150,000 crosses were made between 1975 and 1995 and 20,000 superior genotypes were produced (Fischer and Stapper, 1987). Not only Mexico became self-sufficient in wheat production but India, Pakistan, Turkey and Asian countries were also able to benefit from importing the new varieties and doubling their wheat production also (Ortiz et al., 2007). This essay describes the genes involved in crop dwarfing in wheat and maize.

Semi-dwarf genes – Arabidopsis model

To model the manipulation and effect of these genes, and identify their locations across different crops *Arabidopsis thaliana*. These works were carried out after the numerous wheat crosses with Norin 10 genes. The *gai* mutant allele of *Arabidopsis*, was identified to encode a protein, lacking 17 amino acids from the terminus, which altered the gibberellin (GA) biosynthesis (Peng et al., 1997). Normally, the GAI gene products would repress plant growth in the absence of GA, while they would promote growth in the presence of GA. The mutant repressor, *gai* maintains the repression of GA. As GAs promote plant growth, and the *gai* semi-dominant mutation led to a semi-dwarf plants of heterozygous plants, while in homozygous plants, it led to dwarf varieties. Delayed flowering were observed in short-day photoperiods for these dark green *Arabidopsis* plants.

Peng and Harberd (1993) mapped the *gai* gene region on chromosome 1 using T-DNA insertion in yeast artificial chromosomes (YAC), linked to *GAI* which carried *Dissociation (Ds)* elements. In 1997, Peng et al. continued this work by knocking out the *gai* gene with *Ds* transposons again which led to *gai-t6*, a new allele with the *Ds* insertion. By knocking out *gai*, the plants height was 'tall' again, the same as *GAI* while *gai* led to a dwarf phenotype. This confirmed the function and location of *GAI* and *gai* genes (Figure 2).



Figure 2. Peng et al. (1999) identified the location of *gai* in region I, of the N-terminal region of predicted proteins encoded in comparison to *GAI*.

Gene expression study by Fu et al. (2001) also showed how different levels of *gai* expression leads to different levels of dwarfism in *Arabidopsis*. While molecular work happened in laboratory settings, crop breeding happened in the field and therefore, it is crucial to understand how plant heights and yield changed in the field as well. An extensive study by Würschum et al. (2017) using 1,110 winter wheat variety, showed that from 1950 until 2000, the average wheat height decreased by 25.5 cm (Figure 3). It has also been shown that yield is optimal for 80 cm tall wheat but and it declines for taller varieties (Flintham et al., 1997).

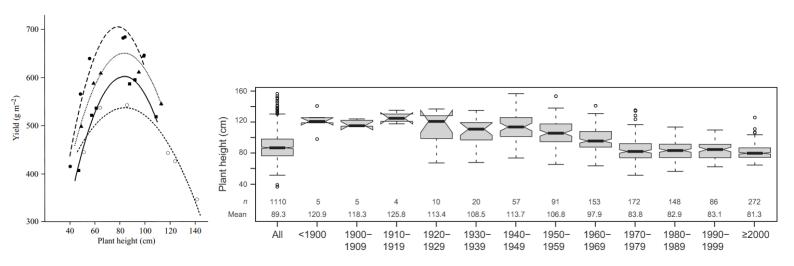


Figure 3. Different *Rht* genotype means (back circles - Maris Huntsman, black squares - Maris Widgeon, black triangles - Bersee, white circle - April Bearded varieties) show a decline in yield after 85 cm plant height (Flintham et al., 1997). Overall height reduction of 1,110 winter wheat from 1900 to 2000 (Würschum et al., 2017)

Wheat Rht genes

The introduction of Norin 10 genes in traditional wheat breeding, formerly called Gai/Rht₁, Gai/Rht₂ genes, started the modern wheat (*Triticum aestivum*) varieties selection to shift towards semi-dwarf phenotypes (Brooking and Kirby, 1981). Twenty-one genes have been described on the wheat's chromosome B and D. *Rht1* to *Rht21* have been described to be responsible for reduced plant height in wheat (McIntosh, 2013). Not all genes were used for crop improvement as some can result in adverse effects of biomass production, decreasing yield.

The four major genes are *Rht-B1b* and *Rht-B1c* on chromosome 4BS, *Rht-D1b* on chromosome and *Rht-D1c* on chromosome 4DS. *Rht-B1b* and *Rht-D1b* encode DELLA proteins which are negative regulators of gibberellins (GA) (Peng et al., 1997; Li et al., 2012). This was identified by Peng et al. (1999; 2002) (Figure 4).



Figure 4. Mutations in the N-terminal regions I (DELLA region) and II (VHYNP region) of wheat *Rht* genes cause GA-insensitivity (Peng et al., 1999).

Rht-B1 was found to contribute 2.7% of the genotypic variance which led to on average 7.6 cm decrease in plant height, and has been identified resistance to Fusarium pathogen. Rht-D1 was found in 49% of the cultivars with the allele substitution effect of -9.4 cm. Rht8 on chromosome 2D, close to Ppd-D1 (photoperiod controller) was identified for future uses as it explained 4.8% genotype variance and led to 9.4 cm height reduction. Rht24 on chromosome 6A, is found in cultivars worldwide and its frequency has been increasing. It was responsible for 15% of genotypic variance on average but the Polish lines showed up to 37.5% while German lines 8.5% genotypic variance.

By gaining an insight to how much variation can be produced by these genes, it enables crop breeding to fine tune these genes and breed selectively to optimise crop reduction to its maximum capacity (Figure 5). These molecular methods have to be tested under field conditions once a new variety is

produced. In Europe now, Community Plant Variety Office (CPVO) and International Union for the Protection of New Varieties of Plants (UPOV), test new varieties by growing them in the field for three years, recording plant characters, combined with molecular work before a new crop variety becomes commercially available.

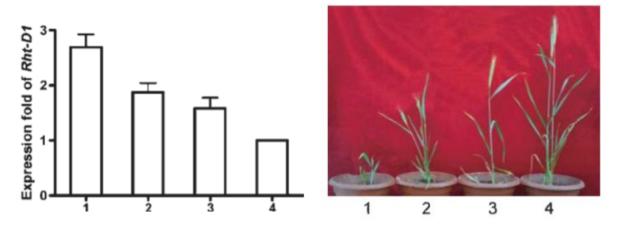


Figure 5. The expression of Rht-D1 expression in transgenic wheat was showed to have the strongest effect on plant height reduction by Li et al. (2012).

Rice modern varieties

At the same time as the modern wheat varieties were developed in 1950s, breeders at the International Rice Research Institute (IRRI) also introduced dwarfing genes into rice (Oryza sativa) plants (Peng et al., 2010). There are two main rice cultivars; indica cultivars require submersion in water and are grown on lowland around tropical Asia and japonica is grown mostly in drier field in temperate East Asia, or at higher altitudes in South Asia. The latter can be temperate or tropical japonica and there are also aromatic, and aus rice cultivars (Wang et al., 2014). The rice dwarfing gene originated from the Chinese indica cultivar, Dee-geo-woo-gen, and was first used used in Taiwanese crosses. In 1950, Taichung Native 1 (TN-1) was produced and in 1966 (Hedden, 2003), IR8 was the new, fertiliser-responsive variety that exceeded the normal yield (6 t/ha⁻¹) to 10 t/ha⁻¹ on irrigated lands (Peng et al., 2010). This was called the 'miracle rice'. The shortened height had the same benefits as in wheat. This TN-1 and IR8 was widely used for semi-dwarf rice indica and japonica cultivars. This was a single recessive allele called sd1 chromosome 1, which was responsible for the semi-dwarf trait. However, addition of GA could counter effect sd1 (Sasaki et al., 2002). This gene has also been shown to control culm length in japonica landraces (Asano et al., 2011). The two genes involved in GA20ox enzyme production, which is involved in GA biosynthesis, were OsGA20ox1 and OsGA20ox2 in rice. The OsGA20ox2 gene originated from the Dee-geo-woo-gen cultivar with a 383-bp deletion in the GA20ox gene (Ashikari et al., 2002). Mutations in these two genes can be used for rice breeding while it was shown that SLR1 gene acts a negative regulator in the GA signalling and therefore leads to a variety of plant heights (Ikeda et al., 2001).

While *sd1* is a gene leading to semi-dwarf phenotype, *d2* is a dwarf gene from *japonica* cultivars (Houghton et al., 2001). A recent study by Yan et al. (2012) gamma radiated KDM105, elite aromatic cultivar from Thailand and produced a *ser* mutant line by a single recessive mutation. This reduced plant height, leaf length, panicle and leaf angle which led to a better performing crop in the field.

Not only the height of the rice plants was selectively bred, but seed shattering was also important; this pattern is controlled by the *sh4* gene (Lin et al., 2007). Tillering capacity is controlled by *moc1* controls in *japonica* cultivar (H89025) while *SCM2* increases spikelet number per panicle in *japonica* (Ookawa et al., 2010).

These improvements led to 132% increase in global rice production from 1966 to 1999 and currently the price of rice is 40% lower than in the 1960s (Khush, 1993).

Maize modern varieties

Maize (Zea mays) dwarf mutants were first described in 1912 by Emerson. Both recessive and dominant alleles are known to cause semi-dwarfism and dwarfism (Harberd and Freeling, 1988). Recessive mutants have been identified on chromosome 3 (d1, d2), d3 on 9S, d5 on 2S and an1 on 1L. Dominant alleles are D8 and Mpl1. The dwarf varieties are reduced in height, and have shorter but broader leaves. The most dominant dwarfing allele was D8-1 described by four amino acid deletion in the DELLA domain, two codon change causing amino acid substitutions.

The mutant alleles of wheat (*Rht*), maize (*d8*) and rice (*sd1*) using the *gai* locations from Peng et al.'s (1999) study, found the N-terminal regions of the *GAI* to be closely related in two regions and was able to locate these regions across the different species (Peng et al., 1999c).

Conclusion

The Green Revolution enabled the human population growth followed by enormous growth of food production systems. However, it has also led to varieties that can take up more fertilisers the use of fertilisers has increased by sevenfold since 1960s (Tilman, 1998). Crop fields are intensive and mostly monocultures which makes them vulnerable to plant pathogens which can lead to total harvest losses (Cleaver, 1972).

Human population is expected to reach 9 billion by 2050 (FAO, 2003) and working towards sustainable food production system is of upmost importance. Climate change is predicted to a 5.8°C temperature rise by the end of the 21st century (Houghton et al., 2001). Over 40% of global food production depends on irrigation systems (Chaves and Oliveira, 2004) and by 2050, 6.5 billion people are predicted to be living in countries facing chronic water shortage (Rockström et al., 2009). Therefore, optimising drought resistance in crops is in the frontline of crop breeding.

Green Revolution could not take place in Africa and some developing countries (Rockström et al., 2007; Toenniessen et al., 2008) without the facilitated help of organisations. Uncovering the functions of genes and fine tuning them with novel molecular techniques to produce optimal harvest quantity and quality with disease resistance (Figure 8), is the future of global food production.

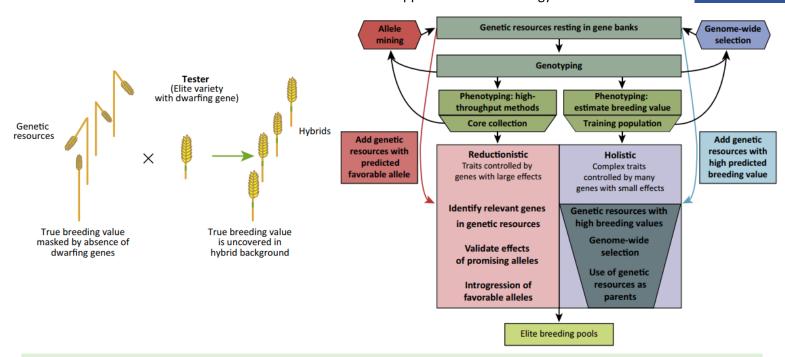


Figure 8. A novel method (left) of wheat variety breeding was proposed (Longin and Reif, 2014) to fully exploit wheat genetic resources by using elite varieties without dwarfing genes, re-introducing dwarfing genes and hybridising the new elite varieties. Selection process can be reductionist or holistic (right), depending if only few traits and genes are focused on or whole genome-wide selection is used.

Word count: 1882

References

Asano, K., Yamasaki, M., Takuno, S., Miura, K., Katagiri, S., Ito, T., Doi, K., Wu, J., Ebana, K., Matsumoto, T. and Innan, H., 2011. Artificial selection for a green revolution gene during *japonica* rice domestication. *Proceedings of the National Academy of Sciences*, 108(27), pp.11034-11039.

Ashikari, M., Sasaki, A., Ueguchi-Tanaka, M., Itoh, H., Nishimura, A., Datta, S., Ishiyama, K., Saito, T., Kobayashi, M., Khush, G.S. and Kitano, H., 2002. Loss-of-function of a rice gibberellin biosynthetic gene, GA20 oxidase (GA20ox-2), led to the rice 'green revolution'. *Breeding Science*, *52*(2), pp.143-150.

Chaves, M.M. and Oliveira, M.M., 2004. Mechanisms underlying plant resilience to water deficits: prospects for water-saving agriculture. *Journal of Experimental Botany*, 55(407), pp.2365-2384

Cleaver, H.M., 1972. The contradictions of the Green Revolution. *The American Economic Review*, 62(1/2), pp.177-186.

Fischer, R.A. and Stapper, M., 1987. Lodging effects on high-yielding crops of irrigated semidwarf wheat. *Field Crops Research*, *17*(3-4), pp.245-258.

Flintham, J.E., Börner, A., Worland, A.J. and Gale, M.D., 1997. Optimizing wheat grain yield: effects of *Rht* (gibberellin-insensitive) dwarfing genes. *The Journal of Agricultural Science*, 128(1), pp.11-25.

Fu, X., Sudhakar, D., Peng, J., Richards, D.E., Christou, P. and Harberd, N.P., 2001. Expression of Arabidopsis *GAI* in transgenic rice represses multiple gibberellin responses. *The Plant Cell*, *13*(8), pp.1791-1802.

Hedden, P., 2003. The genes of the Green Revolution. TRENDS in Genetics, 19(1), pp.5-9.

Hong, Z., Ueguchi-Tanaka, M., Umemura, K., Uozu, S., Fujioka, S., Takatsuto, S., Yoshida, S., Ashikari, M., Kitano, H. and Matsuoka, M., 2003. A rice brassinosteroid-deficient mutant, ebisu dwarf (*d2*), is caused by a loss of function of a new member of cytochrome P450. *The Plant Cell*, *15*(12), pp.2900-2910.

Houghton, J.T., Ding, Y.D.J.G., Griggs, D.J., Noguer, M., van der Linden, P.J., Dai, X., Maskell, K. and Johnson, C.A., 2001. Climate change 2001: The Scientific Basis. The Press Syndicate of the University of Cambridge

Ikeda, A., Ueguchi-Tanaka, M., Sonoda, Y., Kitano, H., Koshioka, M., Futsuhara, Y., Matsuoka, M. and Yamaguchi, J., 2001. *slender* rice, a constitutive gibberellin response mutant, is caused by a null mutation of the *SLR1* gene, an ortholog of the height-regulating gene *GAI/RGA/RHT/D8*. *The Plant Cell*, *13*(5), pp.999-1010.

Khush, G.S., 1993. Breeding rice for sustainable agricultural systems. In *International crop science I. International Crop Science Congress, Iowa, USA, 1992.* (pp. 189-199). Crop Science Society of America

Khush, G.S., 2001. Green revolution: the way forward. Nature Reviews Genetics, 2(10), pp.815-822.

Li, Y., Xiao, J., Wu, J., Duan, J., Liu, Y., Ye, X., Zhang, X., Guo, X., Gu, Y., Zhang, L. and Jia, J., 2012. A tandem segmental duplication (TSD) in green revolution gene *Rht-D1b* region underlies plant height variation. *New Phytologist*, *196*(1), pp.282-291.

Lin, Z., Griffith, M.E., Li, X., Zhu, Z., Tan, L., Fu, Y., Zhang, W., Wang, X., Xie, D. and Sun, C., 2007. Origin of seed shattering in rice (*Oryza sativa L.*). *Planta*, *226*(1), pp.11-20.

Longin, C.F.H. and Reif, J.C., 2014. Redesigning the exploitation of wheat genetic resources. *Trends in plant science*, *19*(10), pp.631-636.

Ortiz, R., Trethowan, R., Ferrara, G.O., Iwanaga, M., Dodds, J.H., Crouch, J.H., Crossa, J. and Braun, H.J., 2007. High yield potential, shuttle breeding, genetic diversity, and a new international wheat improvement strategy. *Euphytica*, *157*(3), pp.365-384.

Peng, J. and Harberd, N.P., 1993. Derivative alleles of the Arabidopsis gibberellin-insensitive (gai) mutation confer a wild-type phenotype. *The Plant Cell*, 5(3), pp.351-360.

Peng, J., Carol, P., Richards, D.E., King, K.E., Cowling, R.J., Murphy, G.P. and Harberd, N.P., 1997. The *Arabidopsis GAI* gene defines a signaling pathway that negatively regulates gibberellin responses. *Genes & development*, *11*(23), pp.3194-3205.

Peng, J., Richards, D.E., Hartley, N.M., Murphy, G.P., Devos, K.M., Flintham, J.E., Beales, J., Fish, L.J., Worland, A.J., Pelica, F. and Sudhakar, D., 1999. 'Green revolution' genes encode mutant gibberellin response modulators. *Nature*, *400*(6741), pp.256-261.

Peng, J., Richards, D.E., Moritz, T., Ezura, H., Carol, P. and Harberd, N.P., 2002. Molecular and physiological characterization of *Arabidopsis GAI* alleles obtained in targeted Ds-tagging experiments. *Planta*, *214*(4), pp.591-596.

Peng, S., Huang, J., Cassman, K.G., Laza, R.C., Visperas, R.M. and Khush, G.S., 2010. The importance of maintenance breeding: A case study of the first miracle rice variety-IR8. *Field crops research*, *119*(2), pp.342-347.

Rockström, J., Falkenmark, M., Karlberg, L., Hoff, H., Rost, S. and Gerten, D., 2009. Future water availability for global food production: the potential of green water for increasing resilience to global change. *Water Resources Research*, *45*(7), W00A12

Rockström, J., Lannerstad, M. and Falkenmark, M., 2007. Assessing the water challenge of a new green revolution in developing countries. *Proceedings of the National Academy of Sciences*, *104*(15), pp.6253-6260.

Sasaki, A., Ashikari, M., Ueguchi-Tanaka, M., Itoh, H., Nishimura, A., Swapan, D., Ishiyama, K., Saito, T., Kobayashi, M., Khush, G.S. and Kitano, H., 2002. Green revolution: a mutant gibberellin-synthesis gene in rice. *Nature*, *416*(6882), pp.701-702.

Tilman, D., 1998. The greening of the green revolution. *Nature*, 396(6708), pp.211-212.

Toenniessen, G., Adesina, A. and DeVries, J., 2008. Building an alliance for a green revolution in Africa. *Annals of the New York academy of sciences*, 1136(1), pp.233-242.

Wang, C.H., Zheng, X.M., Xu, Q., Yuan, X.P., Huang, L., Zhou, H.F., Wei, X.H. and Ge, S., 2014. Genetic diversity and classification of *Oryza sativa* with emphasis on Chinese rice germplasm. *Heredity*, *112*(5), pp.489-496.

Würschum, T., Langer, S.M., Longin, C.F.H., Tucker, M.R. and Leiser, W.L., 2017. A modern Green Revolution gene for reduced height in wheat. *The Plant Journal*, 92, pp.892–903

Yan, W., Hu, B., Zhang, Q., Jia, L., Jackson, A., Pan, X., Huang, B., Yan, Z. and Deren, C., 2012. Short and erect rice (*ser*) mutant from 'Khao Dawk Mali 105' improves plant architecture. *Plant breeding*, *131*(2), pp.282-285.